SEDATIVE ACTION OF SOME SUBSTITUTED BENZYLAMIDES

O.P. SETHI, M.V. GUPTA, O.S. BHATIA AND H.R. DERASARI

Department of Pharmacology and Physiology, L.M. College of Pharmacy, Navrangpura, Ahmedabad-380009

Summary: Substituted benzylamide derivatives of amino acylamide (compound A,B,C, & D) were found to be less potent local anaesthetics than lignocaine and procaine. However, the four compounds exhibited sedation without ptosis and reduced spontaneous locomotor activity better than methaqualone. Compound A alone antagonised methylamphetamine induced hypermotor activity. The test compounds potentiated hexobarbitone induced hypnosis. Three compounds antagonised calcium induced stoppage of isolated heart of frog. Except compound C all caused a transitory fall of blood pressure in dog which was not blocked either by atropine or propranolol. These compounds showed neuromuscular blockade and possessed slight analgesic activity but were devoid of anticonvulsant and tranquillizing activity. LD₅₀ values were calculated to be 164.1 ± 23.0 , 229.1 ± 51.0 , 181.6 ± 28.18 and 416.9 ± 38.2 mg/kg for compounds A,B,C & D respectively.

Key words: basic benzylamides blood pressure fall local anaesthetic neuromuscular blockade CNS actions

calcium antagonist

anticholinergic LD₅₀

INTRODUCTION

Four benzylamides, chemically different from lignocaine, which is an aminoacylamide, were recently synthesised by Parikh (9). All compounds possess lipophilic and hydrophilic chains separated by an intermediate chain. (Fig. 1). Patel and Jindal (10) have shown that

Fig. 1: The chemical structure of the test compounds.

piperidinyl-N-(2:4-diethyl amino N-chlorophenyl) acetamide possessed a potent local anaesthetic activity with low toxicity. Grewal et al. (7) recently reported local anaesthetic action of new substituted acylamide. In the present study, four compounds were subjected to various pharmacological investigations.

MATERIALS AND METHODS

Local anaesthesia:

Conduction, intradermal and surface anaesthesia: The compounds were tested for conduction anaesthesia in mice by the method of Bianchi (1), intradermal anaesthesia on guinea pig back by the method of Bulbring and Wajda (2), and surface anaesthesia on rabbit cornea by the method of Chance and Lobstein (4).

Central nervous system depression:

Spontaneous locomotor and methylamphetamine induced hypermotor activity: Mice weighing between (18-25 g) were used. Motor activity was recorded before and at 1, 2 and 3 hr intervals after the test compounds (5-50 mg/kg i.p.) using 6 beam photoactometer (Techno). Methylamphetamine (4 mg/kg, s.c.) was given and interaction with test compounds was studied. Methaqualone (40 mg/kg, i.p.) was used as a reference drug.

Hexobarbitone hypnosis: Mice weighing (15-25 g) were used. The time of loss and regaining of righting reflex to hexobarbitone (100 and 125 mg/kg, i.p.) was observed before and after the test compounds (10-50 mg/kg, i.p.). Methaqualone (40 mg/kg, i.p.) was used as a reference drug.

Tranquillizing activity:

Conditioned avoidance response (CAR): Rats between (125-250 g) were trained by placing a single rat in Cook's pole climbing apparatus (Techno). Rats were subjected to shock and buzzer at various intervals for 30 sec till the rat climbed the pole immediately. Rats were considered trained if they climbed the pole with the onset of the buzzer. Compounds A,B, C & D (30-50 mg/kg, i.p.) were tested. Chlorpromazine (8 mg/kg, i.p.) was used as a reference drug.

Anticonvulsant activity:

Electrochock and chemoshock convulsions: Mice between (15-25 g) were used. Shocks (60 cycle, 24 mA, and 0.2 sec duration) were delivered through the ear electrodes to mice before and after the compounds (30 to 50 mg/kg, i.p.). Diphenylhydantoin (2.5 to 15 mg/kg) was used as a reference drug. Pentylenetetrazol (80 mg/kg, s.c.) was used before and after the compounds (30-50 mg/kg, i.p.) for chemoshock. Trimethadione (600 mg/kg, i.p.) was used as a reference drug.

Analgesic activity:

Mice tail clip and rat tail flick methods: Albino mice and rats between (18-30 g) and (125-200 g) were used for analgesic activity according to the methods described by Turner (13).

Cardiovascular system:

Isolated heart of frog: Frogs were pithed. Isolated heart of frog was perfused through Cyme's cannula. Responses to adrenaline, potassium chloride, calcium chloride and acetylcholine were elicited before and after the test compounds (10-50 μ g/ml).

Blood pressure of dog: Dogs of either sex were anaesthetised with pentobarbitone (30 mg/kg, i.v.). Blood pressure was recorded with pressure displacement transducer using Encardiorite polygraph from the common carotid artery. The test compounds (6-10 mg/kg) were given after the control panel of adrenaline, noradrenaline, isoprenaline, acetylcholine and histamine. The modifications of responses were recorded.

Skeletal and smooth muscles:

Isolated rectus abdominis muscle of frog and rat ileum: Rectus was mounted in 15 ml organ bath according to the method of Burn (3). Acetylcholine was used before and after the test compounds (15-50 μ g/ml). Rat's ileum was mounted and modifications of acetylcholine responses by the test compounds were recorded.

Acute toxicity :

 LD_{50} estimation: Mice (18-25 g) were given graded doses of the test compounds intraperitoneally. Mice were observed for 24 hrs. LD_{50} values were calculated according to Miller and Tainter method (8).

RESULTS

Local anaesthesia:

Conduction, intradermal and surface anaesthesia: Compounds A. B. C & D showed 30-100% conduction, 30-98% intradermal and 25-85% surface anaesthetic activity respectively. These were less potent than lignocaine and procaine (Table I).

TABLE I: Local anaesthetic activity.

Drugs & compounds		% concentration or nearly 100% activity	% concentration giving 50% or nearly 50% local anaes- thetic activity	
		Intradermal	Conduction	Surface
Lignocaine	40 15	0.3	0.33	0.27*
Procaine		0.5	0.66	
Compound A		1.0	1.50	1.50
Compound B		3.0	4.00	3.00
Compound C		2.0	3.0	2.00
Compound D		1.0	2.0	1.00

^{*}Lignocaine showed 100% surface anaesthetic activity.

Central nervous system depression:

Spontaneous locomotor and methylamphetamine induced hypermotor activity: Compounds A, B, C & D exhibited a dose dependant reduction of locomotor activity recorded in 3 hrs (Table II). Table III summarizes the antagonism of the test compounds A,B,C & D against methylamphetamine and comparison with methaqualone.

TAPLE II : Spontaneous locomotor activity. Locomotor activity recorded for 10 min. (Five mice in each group).

No. of	Drugs & dose (mg/kg, i.p.)	and take topical topic	Count % inhibition after drug					mean %
groups			b	efore.	1 hr	2 hr	3 hr	inhibition in
					1 767	2 111	3 16	Laborated St
2	Normal saline			650	10	20	07	19.7
2	0.1 ml/10 g Methaqualone			630	to about the	20	27	19.7
add and	40			590	60	68	77	68.3
	Compound A	diber to be		330	1 109	00	A shusses	inter test
2	10			490	50	75	. 78	67.7
2	20			530	60	79	85	74.7
2	30			600	78	84	90	84.0
2	Normal saline				de data			
0	0.1 ml/10 g	ill in escol Daby		456	(9.85-8) 1507	27	32	24.7
2	Methaqualone			100	all coll beyon	do mon o	MIN.	portioner.
	Compound B			402	75	78	83	78.7
2	Lompound B			380	36	45	63	48.0
2	30			379	56	69	82	69.0
2	50			341	83	84	82	83.0
2	Normal saline			750	25	34	40	33.0
	0.1 ml/10 g							
2	Methaqualone			tion i	ampolar Mi	* 80-01 m	in Laco	
	40			685	56	72	76	68.0
	Compound C				-	70	EC	53.0
2	5			835	60	73 55	56 44	60.3
2	10			800	82 85	83	84	84.0
2	Normal saline		milital manes	700	20	30	36	28.7
2	Normal saline $0.1 ml/10 g$			700	20	30	30	20.7
2	Methaqualone							
	40			650	58	75	79	70.7
	Compound D							
2	10	te.e	2.0	590	54	74	80	69.3
2	30			568	63	73	87	74.3
' 2	50		0.3	574	76	89	93	86.0
		West Contract	9.0	4 31			1 6	bangana

Methaqualone caused sedation & ptosis. Compounds A,B,C, & D caused sedation alone.

Table III: Methylamphetamine induced hypermotor activity. Locomotor and methylamphetamine (4 mg/kg, subcut) induced hypermotor activity was recorded for 10 min. Each group had 5 mice.

A STATE OF THE PARTY OF THE PAR	and the same of th		Broup nac o mice.					
No. of groups	Drugs	Dose (mg/kg, i.p.)	Cumulative locomotor activity for 10 min (expressed as % of control)	% Inhibition				
2	Normal saline*	0.1 ml/10 g	100					
2	Normal saline	0.1 ml/10 g	198	****				
2	Methaqualone	40	102	****				
2	Compound A	10	138	96				
2		20	133	60				
2		30	110	65				
		30	110	88				
2	Normal saline*	0.1 ml/10 g	100					
2	Normal saline	0.1 ml/10 g	220					
2	Methaqualone	40	105	115				
2	Compound B	10	219	1				
		30	215	5 .				
		50	215	5				
2	Normal saline*	0.1 - 1/10						
2	Normal saline	0.1 ml/10 g	100	****				
2	Methaqualone	0.1 ml/10 g	180	****				
2	Compound C	40	83	97				
Y STATE OF THE STA	Compound C	5	175	5				
		10	168	12				
		30	160	20				
2	Normal saline*	0.1 ml/10 g	100					
2	Normal saline	0.1 ml/10 g	183	****				
2	Methaqualone	40	80	103				
2	Compound D	10	178	5				
		30	175	8				
		50	173	10				
				10				

^{*}Methylamphetamine was not given .

Hexobarbitone hypnosis: Compounds A,B, C & D afforded slight to highly significant potentiation of hexobarbitone hypnosis. Fig. 2 shows comparison with methaqualone.

Tranquillizing activity:

Conditioned avoidance response (CAR): None of the compounds showed any tranquillizing activity, which was evident from CAR study.

Anticonvulsant activity:

Electroshock and chemoshock convulsions: None of the test compounds showed any significant anticonvulsant activity. Diphenylhydantoin and trimethadione afforded complete protection against MES and chemoshock convulsions.

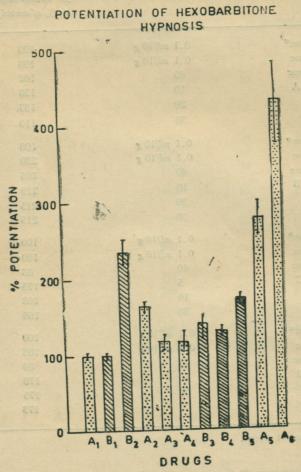


Fig. 2: Potentiation of hexobarbitone hypnosis by the test compounds. A₁: hexobarbitone 100 mg/kg; A₂: hexobarbitone 100 mg/kg + Methaqualone 40 mg/kg; A₃: hexobarbitone 100 mg/kg + compound A 30 mg/kg; A4 : hexobarbitone 100 mg/kg + compound A 50 mg/kg; A_5 : hexobarbitone 100 mg/kg + compound D 30 mg/kg; A_6 : hexobarbitone 100 mg/kg + compound D 50 mg/kg; B₁: hexobarbitone 125 mg/kg: B₂: hexobarbitone 125 mg/kg + Methaqualone 40 mg/kg: B_3 : hexobarbitone 125 mg/kg + compound B 30 mg/kg; B_4 : hexobarbitone 125 mg/kg + compound C 10 mg/kg; B₅: hexobarbitone 125 mg/kg + compound C 30 mg/kg.

Analgesic activity:

Mice tail clip and rat tail flick methods: Compound B exhibited slight analgesic activity, whereas compounds A,C & D were devoid of it.

Cardiovascular system:

Isolated heart of frog: Compounds A,B, C & D showed slight bradycardia in isolated frog heart. Compounds A, C & D antagonised the calcium induced stoppage of heart (Fig. 3), whereas compound B was devoid of it.

ISOLATED FROG HEART

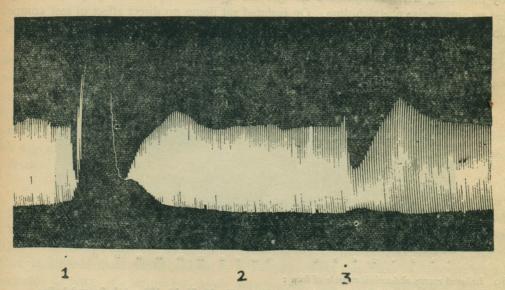


Fig. 3: Isolated heart of frog.
 At 1: calcium chloride 1 mg/ml; 2: compound C 20 μg/ml; 3: calcium chloride 1 mg/ml.

Blood pressure of dog: Compounds A,B & D (6-10 mg/kg) respectively exhibited a transitory fall of blood pressure which was not blocked either by atropine or propranolol (Fig.4).

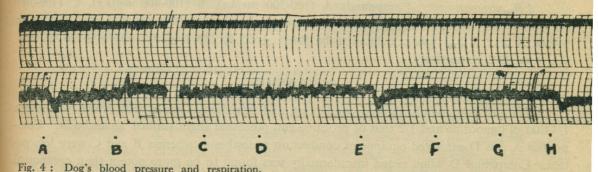


Fig. 4: Dog's blood pressure and respiration.
Upper tracing: Respiration rate
Lower tracing: Blood Pressure At
A: compound D (10 mg/kg); B: Adrenaline (2 μg/kg); C: Atropine suplhate (1 mg/kg); D: Acetylcholine (2 μg/kg); E: compound D (10 mg/kg); F: Propranolol (40 μg/kg); G: Isoprenaline (2 μg/kg) and H: compound D (10 mg/kg).

Compound C did not cause fall of blood pressure. None of the compounds altered the response to adrenaline, noradrenaline, acetylcholine and histamine.

Skeletal and smooth muscles:

Isolated rectus abdominis muscle of frog and rat ileum: Acetylcholine dose response curves were plotted before and in pressence of different doses of the test compounds A,C & D. There was a nonparallel shift of the dose response curve to the right with reduction of the maxima thereby indicating a noncompetitive antagonism against acetylcholine (Fig. 5). Compound B failed to antagonise acetylcholine. On isolated rat ileum no direct effect of test compounds was observed. All compounds $(5-25 \mu g/ml)$ caused reduction of acetylcholine responses.

ISOLATED RECTUS ABDOMINIS MUSCLE OF FROG

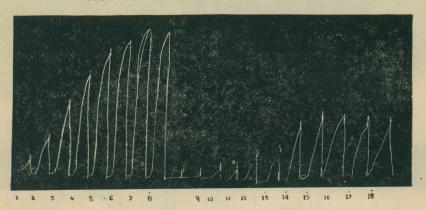


Fig. 5: Isolated rectus abdominis muscle of frog:
At 1, 2, 3, 4, 5, 6, 7, 8 administration of 5, 10, 20, 30, 50, 70, 90, 100 μg/ml of acetylcholine.
At 9, 10, 11, 12, 13, 14, 15, 16, 17, & 18 administration of 5, 10, 10, 50, 100, 200, 300, 400, 600, & 900 μg/ml of acetylcholine, in presence of compound A (10 μg/ml).

Acute toxicity:

 LD_{50} estimations: Compounds A (100-300 mg/kg), B (100-400 mg/kg), C (100-300 mg/kg) and D (300-600 mg/kg) given intraperitoneally showed variable rates of mortality in mice within 24 hrs. LD_{50} was calculated to be 164.1 \pm 23.0, 229.1 \pm 51, 181.6 \pm 28.18 and 416.9 \pm 38.2 respectively.

DISCUSSION

The test compounds showed conduction and intradermal anaesthesia which was about 1/3 that of lignocaine and 1/2 that of procaine and possessed slight surface anaesthesia. Compounds A and D exhibited equipotent conduction anaesthesia whereas B and C were less potent. Compound D when tested intradermally was found to be more potent than compound A, followed by C and B respectively. Compounds B and C possessed the basic structure I.

Compound C (R_2 =CH₃) was more potent as intradermal anaesthetic than compound B (R_2 = C1). The difference in activity could be explained by different substitutions at R_2 . The compound in which aryl group was substituted with electron donating groups (ROR. NR,

R₂, NHCOR, etc) at para-position might be more potent than those in which aryl group was substituted by electron withdrawing groups (RNO, COR, Cl, etc.)

Out of compounds A and C, A was more active than C as it possessed piperidino, whereas compound C possessed morphilino moiety. This observation confirms the finding of Grewal et al. (7) and Weidman et al. (14), who showed that the compounds with piperidino moiety are more potent than those with morphilino moiety as local anaesthetics. Compound D with one aryl group showed the maximum local anaesthetic activity. This could be explained as the optimum balance between the lipophilic and hydrophilic chains is obtained, when there is a single aryl group in the molecule as in compound D than when there are more than one, as in the remaining compounds.

Compounds A, B C & D caused a dose graded reduction in the locomotor activity of the mice whereas compounds A,B & D were more potent than methaqualone. This indicates that the three compounds possess better sedative activity or CNS depressant action than methaqualone. Lignocaine has been reported by Weidling (15) to possess this activity. Compounds B, C & D differed from methaqualone in not antagonising the methylamphetamine induced hypermotor activity. Compound A alone antagonised this. This observation indicates that the compound A may be acting at the sites where methylamphetamine is acting, whereas B, C & D depresses the CNS at other sites. Further, support for its sedative action comes from the potentiation of hexobarbitone hypnosis by all the compounds. Compound D exhibited a highly significant potentiation (4 to 5 times) than methaqualone while others were less effective. Wilder Smith et al. (16) showed potentiation of hexobarbitone induced hypnosis by lignocaine. The present study does not provide evidence to show possible interaction of the test compounds with hexobarbitone metabolism in the liver.

The test compounds were devoid of any tranquillizing or anticonvulsant activity. Compounds A, C & D exhibited a slight analgesic action which is also in accordance with a similar effect of lignocaine.

Compounds A, B, C & D caused slight bradycardia and antagonised calcium induced stoppage of frog heart, probably due to their local anaesthetic activity, Feinstein (6) and Shah et al (11). Compounds A, B & D exhibited a transitory fall of blood pressure in dogs, whereas, compound C did not. This fall may have been due to a direct vasodilator or histamine like action on the blood vessels, as it was not blocked either by atropine or propranalol. Responses to

adrenaline, nor-adrenaline, isoprenaline, acetylcholine and histamine on blood pressure were not modified by these compounds.

Compounds A, C & D showed a dose graded reduction in response to acetylcholine on rectus abdominis muscle of frog which appeared to be of a non-competitive type, indicating a neuromuscular blocking action reported earlier by Derasari et al. (5) and Sinha et al. (12) for local anaesthetics.

Acute toxicity studies showed LD₅₀ values to be 164.1 ± 23.0 , 229.1 ± 51.0 , 181.6 ± 28.8 and 416.4 ± 38.2 for compounds A,B, C & D respectively.

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